ConducTORs of a Signaling Symphony: Metabolic and Hormone Responses Converge on TOR and EIN2 in plants

Jacob O. Brunkard*, Caren Chang, Bruce J. Mayer**, Christian Meyer, Jen Sheen https://doi.org/10.12703/r-01-000008 Published: 2022 May 10

EVALUATION OF

mark EVALUATION BY

The TOR-EIN2 axis mediates nuclear signalling to modulate plant growth.

Fu *et al.* https://doi.org/10.1038/s41586-021-03310-y Article published: 2021 Mar 591:288–92

Development is coordinated by dozens of signals that act in overlapping pathways to orchestrate multicellular growth. Understanding how signaling pathways intersect and diverge at a molecular level is critical to predicting how organisms will react to dynamic environmental conditions. In plants, two antagonistic signaling hubs are strictly required to sense and respond to many nutrients and hormones: TARGET OF RAPAMYCIN (TOR) and ETHYLENE INSENSITIVE 2 (EIN2). In this Landmark report, Fu *et al.* discover that TOR and EIN2 directly interact to choreograph growth and define an unexpected molecular mechanism at the intersection of hormonal and metabolic signaling networks¹.

\land Landmark



Jacob O. Brunkard* University of Wisconsin, Madison Plant genetics & signal transduction



Caren Chang University of Maryland, College Park Ethylene signal transduction



Bruce J. Mayer** University of Connecticut School of

Medicine Cell signaling mechanisms



Christian Meyer

IJPB, INRAE, Université Paris-Saclay TOR kinase and nitrogen metabolism



Harvard Medical School; Massachusetts General Hospital *Plant signaling*

*Corresponding author and primarily responsible for drafting the consensus evaluation: Jacob O. Brunkard (brunkard@wisc.edu)

****Broad-perspective** panelist from a different field

Competing interests: The authors declare that they have no competing interests.

Copyright: 2022 Faculty Opinions Ltd.

How to cite this article: Brunkard et al. ConducTORs of a Signaling Symphony: Metabolic and Hormone Responses Converge on TOR and EIN2 in plants. Fac Rev 2022, 11:(12) (https://doi.org/10.12703/r-01-000008)

Articles evaluated for landmarks are selected by members of an Advisory Board.

Background

Eukaryotes coordinate metabolism through a conserved signaling hub, the TARGET OF RAPAMYCIN (TOR) atypical serine/threonine kinase^{2–9}. Nutrients and some hormones stimulate TOR, which phosphorylates substrate proteins to engage multiple downstream pathways that broadly promote growth and anabolism¹⁰⁻¹³. When nutrients are limiting or conditions are unfavorable for growth, TOR activity declines and cells become quiescent¹⁴. TOR dysregulation causes or contributes to diverse human diseases, including cancers and age-related disorders, which has provoked significant investigation of TOR signaling networks in biomedical models¹⁵. These efforts have only recently started to reveal how TOR can decipher myriad upstream cues to modulate precise downstream responses. Much less is known about the TOR signaling network in plants, but plant biologists are increasingly interested in the potential benefits of genetically leveraging TOR regulatory systems to create resilient, high-yielding crops for a sustainable agricultural future.

Plants continually grow during their vegetative life cycle through cell division and expansion at shoot and root meristems and through cell expansion beyond the meristems¹⁶. Cell division and expansion are both developmentally coordinated by several phytohormones, including the gaseous phytohormone ethylene. Ethylene is popularly familiar for its role in fruit ripening: many fruits, such as apples and bananas, depend on ethylene for ripening. Ethylene also promotes seed germination, regulates development, and mediates responses to various abiotic and biotic stresses. At a molecular level, ethylene engages a well-defined signal transduction cascade that was first dissected through forward genetic screens for ethylene-insensitive mutants of Arabidopsis thaliana (Figure 1)¹⁷⁻¹⁹. One of the ethylene signaling components, ETHYLENE-INSENSITIVE 2 (EIN2), is a sig-

naling hub of elusive molecular function. EIN2 appears to play multifaceted roles in plant cells, since several *ein2* alleles have been found in forward genetic screens for responses to various signals, including glucose²⁰, paraquat-triggered oxidative stress²¹, and the phytohormones auxin²², cytokinin²³, abscisic acid^{24,25}, and jasmonic acid²¹. Since ethylene responses are blocked in ein2 mutants, these effects could reveal general connections between ethylene biosynthesis/signaling and other pathways (e.g., cytokinin acts, in part, by promoting ethylene biosynthesis), but several of these responses cannot be readily explained through the role of EIN2 in ethylene signaling. In their Landmark report, Fu et al.¹ make major advances in understanding how TOR and EIN2, two molecular signaling hubs, cooperate to coordinate plant responses to diverse upstream cues and regulate growth and development.

Main contributions and importance

When seedlings are grown in complete darkness, a situation analogous to germination under the soil, their hypocotyls (embryonic stems) elongate until the seedlings encounter light. Unlike many stages of plant development, hypocotyls elongate exclusively through cell expansion, not division²⁶⁻²⁸. TOR and ethylene antagonistically regulate hypocotyl elongation: ethylene causes dark-grown seedlings to form short, thick hypocotyls¹⁹, whereas TOR promotes long, narrow hypocotyl growth²⁹. To discover how TOR promotes hypocotyl elongation, Fu et al. screened for mutants involved in hypocotyl growth and discovered that mutants defective in ethylene responses, ein2 and ein3; ein3-like1, are less sensitive to TOR inhibition and continue to grow even when TOR is inactivated. Surprisingly, however, etr1 mutants defective in ethylene sensing upstream of EIN2 are not resistant to TOR inhibition, ethylene does not impact TOR activity in dark-grown seedlings, and inhibitors of ethylene biosynthesis and signaling also had no impact on TOR regulation of hypocotyl

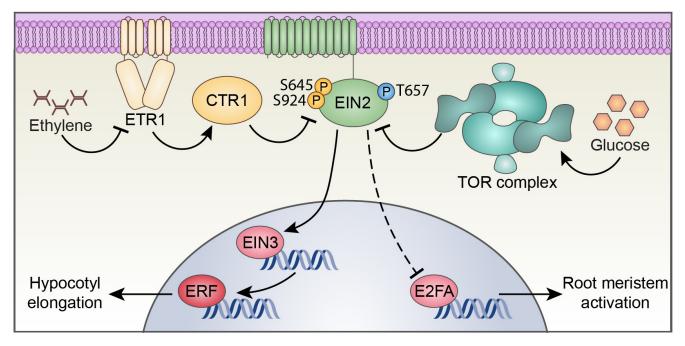


Figure 1. Ethylene-ETR1-CTR1 and glucose-TOR signaling converge on the EIN2 signaling hub

Ethylene is perceived at the ER membrane (purple) by a family of receptors that includes ETR1^{30,31}. In the absence of ethylene, the receptors activate a cytosolic serine/threonine kinase, CTR1, which phosphorylates another ER-resident protein, EIN2, at residues Ser645 and Ser924³¹. Ethylene directly binds to and suppresses ETR1 receptors, preventing their activation of CTR1, and EIN2 (in the unphosphorylated state) is then proteolytically cleaved to release its cytosolic C-terminus from the ER membrane^{32–34}; this fragment ("EIN2-C") promotes the expression and activity of EIN3 family transcription factors through multiple mechanisms in the cytosol and nucleus (blue)^{35–39}. EIN3 transcription factors drive the ethylene-response transcriptional program, including by directly promoting transcription of the much larger ERF family of ethylene-response transcription factors^{40–42}. A classical consequence of ethylene signaling is repressed hypocotyl elongation in the dark¹⁹. In this Landmark study, Fu *et al.* discover that the TOR complex phosphorylates another EIN2 residue, Thr657. They present evidence that full-length EIN2, which potentially has distinct functions from EIN2-C²¹, accumulates in the nucleus when Thr657 is not phosphorylated by TOR¹. When metabolic conditions are not favorable and TOR is inactive, Thr657 is not phosphorylated and EIN2 represses root meristem activity, at least in part by preventing E2FA-promoted cell cycle progression¹.

elongation. These results strongly suggested that TOR acts through an ethylene-independent pathway that converges on EIN2 to coordinate growth.

Fu *et al.* next deployed a series of elegant experiments to demonstrate that TOR directly interacts with EIN2 and phosphorylates Thr657, a broadly conserved residue in orthologues of EIN2 that had not been previously investigated. Biochemically mimicking EIN2-Thr657 phosphorylation in mutated "phosphomimetic" EIN2-T657D transgenic lines was sufficient to render seedlings insensitive to TOR inhibition, demonstrating that TOR-EIN2 signaling is critical to drive hypocotyl elongation. Strikingly, the phosphomimetic EIN2-T657D lines remain fully sensitive to ethylene, which prevents phosphorylation of two different EIN2 residues, Ser645 and Ser924. Oppositely, phosphomimetic EIN2-S645D lines are ethylene-insensitive but remain sensitive to TOR inhibitors. Therefore, TOR and ethylene signaling intersect at the EIN2 signaling hub but act on EIN2 through distinct phosphosites to regulate hypocotyl elongation. Moreover, transcriptional analysis of light-grown wild-type and mutant seedlings revealed that EIN2 is required for a majority of the glucose-triggered, TOR-dependent responses, suggesting that EIN2 is a key effector of TOR metabolic programming in plants.

Open questions

This Landmark study provides a compelling model for understanding how cellular signal transduction networks interconnect and opens several new avenues for investigation in plant biology. Fu *et al.* provide evidence that TOR-catalyzed EIN2-Thr657 phosphorylation prevents translocation of full-length EIN2 from the ER to the nucleus, whereas ethylene promotes cleavage and release of a soluble C-terminal fragment of EIN2 (EIN2-C) that promotes ethylene responses in the nucleus and cytosol. This raises the possibility that full-length EIN2 has distinct activities from EIN2-C in the nucleus, and might constitute an uncharacterized translocation mechanism, since it is not obvious how a transmembrane ER protein could relocalize to the nucleus.

At the organismal level, both ethylene and TOR regulate growth, development, and physiology in contexts beyond the Arabidopsis seedling models used in this study. Does EIN2 mediate TOR signaling in these contexts? How do TOR and EIN2 interact to modulate responses to metabolic status, ethylene, and other phytohormones when plants experience abiotic stress, during ripening and senescence, or during pathogen attack? The discovery that TOR and EIN2 work closely together could help to illuminate how phytohormone signals and metabolic cues intersect throughout a plant's lifespan.

Evolutionarily, ethylene signaling arose in early algal ancestors of plants⁴³ and TOR was already present in the last eukaryotic common ancestor⁹. Moreover, the TOR-catalyzed phosphosite of EIN2, Thr657, appears to be conserved even in some bryophytes, hinting that TOR-EIN2 regulation may have evolved in the earliest land plants. Therefore, determining whether the TOR-EIN2 signaling hub is functionally conserved beyond the Arabidopsis model system could reveal new targets for agricultural scientists working to breed resilient, high-yielding crops.

Conclusion

The convergence of TOR and EIN2 signaling networks through direct molecular and functional interactions illustrates how complex upstream cues can be deciphered by cells to modulate specific downstream responses. This creative investigation from Fu *et al.* is a stellar example of how cell and molecular biology can be used to address classical problems—in this case, how plants integrate various signals, including nutrients and hormones, to coordinate growth—and reveal underlying mechanisms. Going forward, the discovery of the TOR-EIN2 signaling hub will serve as a model for investigations of cellular signal transduction and provoke new fundamental and translational research in plant physiology and development.

References

1. 🐼 Landmark

Fu L, Liu Y, Qin G, *et al.* 2021. **The TOR-EIN2 axis mediates nuclear signalling to modulate plant growth.** *Nature* **591**:288–92. doi: 10.1038/s41586-021-03310-y

- Kunz J, Henriquez R, Schneider U, et al. 1993.
 Target of rapamycin in yeast, TOR2, is an essential phosphatidylinositol kinase homolog required for G₁ progression. Cell 73:585–96. doi: 10.1016/0092-8674(93)90144-F
- Cafferkey R, Young PR, McLaughlin MM, et al. 1993.
 Dominant missense mutations in a novel yeast protein related to mammalian phosphatidylinositol 3-kinase and VPS34 abrogate rapamycin cytotoxicity. Mol Cell Biol 13:6012–23. doi: 10.1128/mcb.13.10.6012-6023.1993
- Brown EJ, Albers MW, Shin TB, et al. 1994. A mammalian protein targeted by G1-arresting rapamycin-receptor complex. *Nature* 369:756–8. doi: 10.1038/369756a0
- Sabatini DM, Erdjument-Bromage H, Lui M, et al. 1994. RAFT1: A mammalian protein that binds to FKBP12 in a rapamycin-dependent fashion and is homologous to yeast TORs. Cell 78:35–43. doi: 10.1016/0092-8674(94)90570-3
- 6. Recommended
 Menand B, Desnos T, Nussaume L, et al. 2002.
 Expression and disruption of the Arabidopsis TOR (target of rapamycin) gene. Proc Natl Acad Sci U S A 99:6422–7.
 doi: 10.1073/pnas.092141899

doi. 10.1073/pilas.092141899

 Blenis J. 2017. TOR, the Gateway to Cellular Metabolism, Cell Growth, and Disease. Cell 171:10–3. doi: 10.1016/j.cell.2017.08.019

- Valvezan AJ, Manning BD. 2019. Molecular logic of mTORC1 signalling as a metabolic rheostat. Nat Metab 1:321–33. doi: 10.1038/s42255-019-0038-7
- Brunkard JO. 2020. Exaptive Evolution of Target of Rapamycin Signaling in Multicellular Eukaryotes. *Dev Cell* 54:142–55. doi: 10.1016/j.devcel.2020.06.022
- Melick CH, Jewell JL. 2020. Regulation of mTORC1 by Upstream Stimuli. *Genes (Basel)* 11:989. doi: 10.3390/genes11090989
- Li L, Liu KH, Sheen J. 2021. Dynamic Nutrient Signaling Networks in Plants. Annu Rev Cell Dev Biol 37:341–67. doi: 10.1146/annurev-cellbio-010521-015047

12. 🔊 Recommended

van Leene J, Han C, Gadeyne A, *et al.* 2019. **Capturing the phosphorylation and protein interaction landscape of the plant TOR kinase.** *Nat Plants* **5**:316–27. doi: 10.1038/s41477-019-0378-z

 Scarpin MR, Leiboff S, Brunkard JO. 2020. Parallel global profiling of plant TOR dynamics reveals a conserved role for LARP1 in translation. *eLife* 9: e58795. doi: 10.7554/eLife.58795

14. 🔊 Recommended

Xiong Y, McCormack M, Li L, *et al.* 2013. **Glucose-TOR** signalling reprograms the transcriptome and activates meristems. *Nature* **496**:181–6. doi: 10.1038/nature12030

dol: 10.1038/nature1203

15. 🔊 Recommended

Liu GY, Sabatini DM. 2020. **mTOR at the nexus of nutrition, growth, ageing and disease.** *Nat Rev Mol Cell Biol* **21**:183–203. doi: 10.1038/s41580-019-0199-y 16. Sluis A, Hake S. 2015. Organogenesis in plants: Initiation and elaboration of leaves. Trends Genet 31:300-6. doi: 10.1016/j.tig.2015.04.004

- 17. Roman G, Lubarsky B, Kieber JJ, et al. 1995. Genetic analysis of ethylene signal transduction in Arabidopsis thaliana: Five novel mutant loci integrated into a stress response pathway. Genetics 139:1393-409. doi: 10.1093/genetics/139.3.1393
- 18. Binder BM. 2020. Ethylene signaling in plants. J Biol Chem 295:7710-25. doi: 10.1074/jbc.REV120.010854
- 19. Bleecker AB, Estelle MA, Somerville C, et al. 1988. Insensitivity to Ethylene Conferred by a Dominant Mutation in Arabidopsis thaliana. Science 241:1086-9. doi: 10.1126/science.241.4869.1086
- 20. Recommended

Yanagisawa S, Yoo SD, Sheen J. 2003. Differential regulation of EIN3 stability by glucose and ethylene signalling in plants. Nature 425:521-5. doi: 10.1038/nature01984

- 21. Alonso JM, Hirayama T, Roman G, et al. 1999. EIN2, a bifunctional transducer of ethylene and stress responses in Arabidopsis. Science 284:2148-52. doi: 10.1126/science.284.5423.2148
- 22. Fujita H, Syono K. 1996. Genetic analysis of the effects of polar auxin transport inhibitors on root growth in Arabidopsis thaliana. Plant Cell Physiol 37:1094–101. doi: 10.1093/oxfordjournals.pcp.a029059
- 23. Cary AJ, Liu W, Howell SH, et al. 1995. Cytokinin action is coupled to ethylene in its effects on the inhibition of root and hypocotyl elongation in Arabidopsis thaliana seedlings. Plant Physiol 107:1075-82. doi: 10.1104/pp.107.4.1075
- 24. Cutler S, Ghassemian M, Bonetta D, et al. 1996. A protein farnesyl transferase involved in abscisic acid signal transduction in Arabidopsis. Science 273:1239-41. doi: 10.1126/science.273.5279.1239
- 25. Beaudoin N, Serizet C, Gosti F, et al. 2000. Interactions

between abscisic acid and ethylene signaling cascades. Plant Cell 12:1103-15. doi: 10.1105/tpc.12.7.1103

- 26. Gendreau E, Traas J, Desnos T, et al. 1997. Cellular basis of hypocotyl growth in Arabidopsis thaliana. Plant Physiol 114:295-305. doi: 10.1104/pp.114.1.295
- 27. Raz V, Koornneef M. 2001. Cell division activity during apical hook development. Plant Physiol 125:219-26. doi: 10.1104/pp.125.1.219
- 28. Kutschera U, Niklas KJ. 2013. Cell division and turgordriven stem elongation in juvenile plants: A synthesis. Plant Sci 207:45-56. doi: 10.1016/j.plantsci.2013.02.004

29. 🔊 Recommended

Ren M, Venglat P, Qiu S, et al. 2012. Target of rapamycin signaling regulates metabolism, growth, and life span in Arabidopsis. Plant Cell 24:4850-74. doi: 10.1105/tpc.112.107144

- 30. Chang C, Kwok SF, Bleecker AB, et al. 1993. Arabidopsis ethylene-response gene ETR1: Similarity of product to two-component regulators. Science 262:539-44. doi: 10.1126/science.8211181
- 31. Schaller GE, Bleecker AB. 1995. Ethylene-binding sites generated in yeast expressing the Arabidopsis ETR1 gene. Science 270:1809–11. doi: 10.1126/science.270.5243.1809

32. 🔊 Recommended

Ju C, Yoon GM, Shemansky JM, et al. 2012. CTR1 phosphorylates the central regulator EIN2 to control ethylene hormone signaling from the ER membrane to the nucleus in Arabidopsis. Proc Natl Acad Sci USA **109**:19486–91. doi: 10.1073/pnas.1214848109

33. 🔊 Recommended

Qiao H, Shen Z, Huang SC, et al. 2012. Processing and subcellular trafficking of ER-tethered EIN2 control response to ethylene gas. Science 338:390-3. doi: 10.1126/science.1225974

34. 🔊 Recommended

Wen X, Zhang C, Ji Y, *et al.* 2012. Activation of ethylene signaling is mediated by nuclear translocation of the cleaved EIN2 carboxyl terminus. *Cell Res* 22:1613–6. doi: 10.1038/cr.2012.145

35. 🔊 Recommended

Merchante C, Brumos J, Yun J, *et al.* 2015. Gene-specific translation regulation mediated by the hormone-signaling molecule EIN2. *Cell* 163:684–97. doi: 10.1016/j.cell.2015.09.036

36. 🔊 Recommended

Li W, Ma M, Feng Y, *et al.* 2015. **EIN2-directed translational regulation of ethylene signaling in** *Arabidopsis. Cell* **163**:670–83. doi: 10.1016/j.cell.2015.09.037

37. 🐼 Recommended

Zhang F, Qi B, Wang L, *et al.* 2016. **EIN2-dependent** regulation of acetylation of histone H3K14 and noncanonical histone H3K23 in ethylene signalling. *Nat Commun* 7:13018. doi: 10.1038/ncomms13018

 Zhang F, Wang L, Qi B, et al. 2017. EIN2 mediates direct regulation of histone acetylation in the ethylene response. Proc Natl Acad Sci USA 114:10274–9. doi: 10.1073/pnas.1707937114

- Wang L, Zhang Z, Zhang F, et al. 2021. EIN2-directed histone acetylation requires EIN3-mediated positive feedback regulation in response to ethylene. *Plant Cell* 33:322–37. doi: 10.1093/plcell/koaa029
- 40. Chao Q, Rothenberg M, Solano R, et al. 1997. Activation of the Ethylene Gas Response Pathway in Arabidopsis by the Nuclear Protein ETHYLENE-INSENSITIVE3 and Related Proteins. Cell 89:1133–44. doi: 10.1016/S0092-8674(00)80300-1
- 41. Solano R, Stepanova A, Chao Q, et al. 1998. Nuclear events in ethylene signaling: A transcriptional cascade mediated by ETHYLENE-INSENSITIVE3 and ETHYLENE-RESPONSE-FACTOR1. Genes Dev 12:3703–14. doi: 10.1101/gad.12.23.3703
- Ohme-Takagi M, Shinshi H. 1995. Ethylene-inducible DNA binding proteins that interact with an ethyleneresponsive element. *Plant Cell* 7:173–82. doi: 10.1105/tpc.7.2.173

43. 🔊 Recommended

Ju C, van de Poel B, Cooper ED, *et al.* 2015. **Conservation of ethylene as a plant hormone over 450 million years of evolution.** *Nat Plants* 1:14004. doi: 10.1038/nplants.2014.4